



# The Louisiana Animal Disease Diagnostic Laboratory

A service unit of the LSU School of Veterinary Medicine

Adapted from Murphy, F.A., et al, Veterinary Virology, 3rd ed.  
Academic Press, 1999. Compiled by Rob Poston

## Sampling and Testing Guide for Common Viral Diseases of Animals in Louisiana

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[Deer and Small Ruminants](#)

[Swine](#)

[Horses](#)

[Dogs](#)

[Cats](#)

[Multi-species: Rabiesvirus](#)

# Cattle

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[Respiratory viral diseases](#)

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# Deer and Small Ruminants

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# Cats

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# Generalized Viral Diseases of Cattle

Please click on the virus of interest:

[Bovine Viral Diarrhea \(BVD\) Virus](#)

[Bovine Herpesvirus Type 1 \(BHV-1, IBR, IPV\)](#)

[Bovine Leukemia Virus \(BLV\)](#)

[Bovine Immunodeficiency Virus \(BIV\)](#)

[Foot and Mouth Disease \(FMD\) virus](#)



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# Respiratory Viral Diseases of Cattle

Please click on the virus of interest:

[Bovine Viral Diarrhea \(BVD\) Virus](#)

[Bovine Herpesvirus Type 1 \(BHV-1, IBR, IPV\)](#)

[Bovine Respiratory Syncytial Virus \(BRSV\)](#)

[Parainfluenza Type 3 \(PI-3\) Virus](#)

[Bovine Respiratory Coronavirus \(BRCV\)](#)

[Bovine Rhinovirus](#)

[Bovine Adenovirus](#)



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# Enteric Viral Diseases of Cattle

Please click on the virus of interest:

[Mucosal Disease](#)

[Bovine Rotavirus \(BRotV\)](#)

[Bovine Enteric Coronavirus \(BECV\)](#)

[Astrovirus, Calicivirus, Torovirus](#)



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# Reproductive/neonatal Viral Diseases of Cattle

Please click on the virus of interest:

[Bovine Viral Diarrhea \(BVD\) Virus](#)

[Bovine Herpesvirus Type 1 \(BHV-1 or IBR\)](#)

[Bluetongue Virus \(BTV\)](#)



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# Viral Infections Affecting the Skin of Cattle

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[Mucosal Disease \(MD\)](#)

[Bovine Viral Diarrhea \(BVD\) Virus](#)

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[Bovine Papilloma Virus](#)

[Vesicular Stomatitis Virus \(VSV\)](#)

[Pseudocowpox Virus](#)

[Foot and Mouth Disease \(FMD\) Virus](#)



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# Generalized Viral Diseases of Small Ruminants

Please click on the virus of interest:

[Bluetongue Virus \(BTV\)](#)

[Epizootic Hemorrhagic Disease \(EHD\) Virus](#)

[Caprine Arthritis-encephalitis \(CAE\) Virus](#)

[Ovine Progressive Pneumonia \(OPP\) Virus](#)

[Maedi / Visna virus](#)

[Malignant Catarrhal Fever \(MCF\) Viruses](#)



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# Respiratory Viral Disease of Small Ruminants

Please click on the virus of interest:

[Parainfluenza 3 \(PI-3\) Virus](#)

[Ovine Adenovirus](#)

[Respiratory Syncytial Virus \(RSV\)](#)

[Ovine Pulmonary Adenomatosis Virus](#)



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# Enteric Viral Disease of Small Ruminants

Please click on the virus of interest:

[Rotavirus](#)

[Coronavirus](#)

[Adenovirus](#)

[Astrovirus](#)



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# Reproductive Viral Diseases in Small Ruminants

Please click on the virus of interest:

[Bluetongue Virus \(BTV\)](#)



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# Viral Skin Diseases of Small Ruminants

Please click on the virus of interest:

[Orf \(Contagious Pustular Dermatitis\)](#)

[Vesicular Stomatitis Virus](#)

[Foot and Mouth Disease \(FMD\) Virus](#)



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# Generalized Viral Diseases of Swine

Please click on the virus of interest:

[Pseudorabies Virus \(PRV\)](#)

[Porcine Reproductive and Respiratory Syndrome \(PRRS\) Virus](#)

[Porcine Hemagglutinating Encephalitis Virus \(HEV\)](#)

[Porcine Circovirus Type 2 \(PCV-2\), Postweaning Multisystemic Wasting Syndrome \(PMWS\)](#)

[Porcine Lymphosarcoma Virus](#)

[Encephalomyelocarditis Virus \(EMCV\)](#)

[Vesicular Stomatitis Virus \(VSV\)](#)

[Hog Cholera \(HC\)](#)



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# Respiratory Viral Disease of Swine

Please click on the virus of interest:

[Pseudorabies Virus \(PRV\)](#)

[Porcine Reproductive and Respiratory Syndrome \(PRRS\) Virus](#)

[Swine Influenza Virus \(SIV\)](#)

[Swine Cytomegalovirus](#)



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# Enteric Viral Disease of Swine

Please click on the virus of interest:

[Porcine Rotavirus \(PRotV\)](#)

[Transmissible Gastroenteritis \(TGE\) Virus](#)



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# Reproductive/neonatal Viral Diseases of Swine

Please click on the virus of interest:

[Pseudorabies Virus \(PRV\)](#)

[Porcine Reproductive and Respiratory Syndrome \(PRRS\) Virus](#)

[Porcine Parvovirus \(PPV\)](#)

[Porcine Enteroviruses 2-11 \(SMEDI group\)](#)



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# Viral Infections Affecting the Skin of Swine

Please click on the virus of interest:

[Vesicular Stomatitis Virus \(VSV\)](#)

[Swinepox Virus](#)

[Foot and Mouth Disease \(FMD\) Virus](#)

[Swine Vesicular Disease](#)

[Vesicular Exanthema of Swine](#)



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# Generalized Viral Diseases of Horses

Please click on the virus of interest:

[Equine Infectious Anemia \(EIA\) Virus](#)

[Equine Viral Arteritis \(EVA\) Virus](#)



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# Neurological Viral Diseases of Horses

Please click on the virus of interest:

[Eastern Equine Encephalitis \(EEE\)](#)

[Venezuelan Equine Encephalitis \(VEE\)](#)

[Western Equine Encephalitis \(WEE\)](#)

[West Nile Viral \(WNV\) Encephalitis](#)

[Rabiesvirus](#)



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# Respiratory Viral Disease of Horses

Please click on the virus of interest:

[Equine Herpesvirus Type-1 \(EHV-1\)](#)

[Equine Herpesvirus Type-4 \(EHV-4\)](#)

[Equine Rhinopneumonitis Virus \(ERV\)](#)

[Equine Influenza Virus \(EIV\)](#)

[Equine Herpesvirus Type 2](#)

[Equine Rhinovirus](#)

[Equine Adenovirus](#)



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# Enteric Viral Disease of Horses

Please click on the virus of interest:

[Rotavirus \(RotV\)](#)

[Corona virus](#)



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# Abortifacient/neonatal Equine Viral Diseases

Please click on the virus of interest:

[Equine Herpesvirus Type-1 \(EHV-1\)](#)  
[Equine Abortion Virus](#)

[Equine Viral Arteritis \(EVA\)](#)



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# Viral Infections Affecting the Skin of Horses

Please click on the virus of interest:

[Vesicular Stomatitis Virus \(VSV\)](#)

[Equine Papilloma Virus](#)

[Equine Sarcoid](#)



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# Generalized Viral Disease of Dogs

Please click on the virus of interest:

[Canine Distemper Virus \(CDV\)](#)

[Canine Parvovirus Virus Type 2 \(CPV\)](#)

[Canine Adenovirus Type 1 \(CAdV-1\)](#)  
[Infectious Canine Hepatitis \(ICH\) virus](#)



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# Respiratory Viral Disease of Dogs

Please click on the virus of interest:

[Canine Distemper Virus \(CDV\)](#)

[Canine Adenovirus Type 2 \(CAAdV-2\)](#)

[Canine Tracheobronchitis Virus](#)

[Canine Parainfluenza Virus \(CPIV\)](#)

[Parainfluenza Virus Type 2 \(PI-2\)](#)

[Canine Herpesvirus \(CHV\)](#)



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# Enteric Viral Disease of Dogs

Please click on the virus of interest:

[Canine Coronavirus \(CCV\)](#)

[Canine Parvovirus Virus Type 2 \(CPV-2\)](#)

[Rotavirus \(RotV\)](#)



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# Reproductive/neonatal Viral Diseases of Dogs

Please click on the virus of interest:

[Canine Herpesvirus \(CHV\)](#)



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# Generalized Viral Diseases of Cats

Please click on the virus of interest:

[Feline Infectious Peritonitis \(FIP\) Virus](#)

[Feline Leukemia/sarcoma Virus \(FeLV\)](#)

[Feline Immunodeficiency Virus \(FIV\)](#)

[Feline Parvovirus \(FPV\)](#)

[Feline Panleukopenia Virus \(FPL\)](#)



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# Respiratory Viral Diseases of Cats

Please click on the virus of interest:

[Feline Herpesvirus \(FHV\)](#)

[Feline Viral Rhinotracheitis \(FVR\)](#)

[Feline Calicivirus \(FCV\)](#)



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# Enteric Viral Disease of Cats

Please click on the virus of interest:

[Feline Parvovirus \(FPV\)](#)

[Feline Panleukopenia Virus \(FPL\)](#)

[Feline Coronavirus \(FECV\)](#)

[Rotavirus \(RotV\)](#)



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# Reproductive/neonatal Diseases of Cats

Please click on the virus of interest:

[Feline Parvovirus \(FPV\)](#)

[Feline Panleukopenia Virus \(FPL\)](#)



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# Guide Key

## Virus name, synonyms (*Family name, subfamily or genus in italics*)

- Systemic or local infection / with or without viral persistence / “major”, “moderate”, “minor”, or “slight” disease significance (minor or slight—testing probably not cost effective until other common differentials considered, relative to the target species population in Louisiana)
- Antemortem or postmortem specimens:
  - Preferred samples; swabs, fluids, tissues or organs (popular technology in parentheses)
  - Viral detection technique abbreviations:
    - EM—Electron microscopy
    - ELISAg—Enzyme-linked antibody assay for viral antigen
    - FA—Fluorescent antibody
    - IHC—Immunohistochemistry
    - ISH—In-situ hybridization
    - PCR—Polymerase chain reaction or similar genetic probe technique
    - HA—Hemagglutination
    - VI—Virus isolation by culture.
  - Serological technique abbreviations:
    - AGID—Agar gel immunodiffusion
    - ELISAb—Enzyme-linked antibody assay for viral antibody
    - HI—Hemagglutination inhibition
    - IFA—indirect fluorescent antibody
    - LA—Latex agglutination
    - PRNT—Plaque reduction neutralization
    - SN—Serum neutralization
  - Some indicated technologies are not currently available at LaVMDL
  - If testing is “not routine”, viral detection technology is not used in diagnosis generally, or may not be readily available; specimens may be sent elsewhere for testing, if necessary.
- Clinicopathology—system involvement, signs, lesions, and possible known consequences of viral infection (relative to species), pathogenesis, natural host range, disease prevalence
- Other relevant diagnostic and epidemiological information, such as serotypes and sub-types, cross-species infections, zoonotic potential



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# Bovine Viral Diarrhea (BVD) Virus (*Flaviviridae, pestivirus*)

- Systemic infection / persistence / major significance
- Antemortem
  - Nasopharyngeal swab or fluid, stool, whole blood; carrier calf's serum (VI, PCR)
  - Nasopharyngeal swab or fluid, serum (ELISA<sub>Ag</sub> kit)
  - Paired serum (Serology—SN, ELISA<sub>Ab</sub>, IFA) (Carrier's serum is virus positive, antibody negative)
- Postmortem
  - Turbinate, trachea, lung, tonsil, ileocecal Peyer's patches, mandibular and mesenteric lymph nodes, cerebellum; fetal lung, liver, kidney, spleen, adrenal, brain (generally unrewarding samples include spleen, thymus, and mucosa of esophagus, rumen, and oral cavity) (FA, VI, PCR)
- Clinicopathology—respiratory, enteric, dermal, mucosal disease, reproductive (early embryonic death, abortion), runting, cerebellar hypoplasia, and persistently infected carrier calves
- BVDV also causes Mucosal Disease



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# Bovine Herpesvirus Type 1 (BHV-1)

## Infectious Bovine Rhinotracheitis (IBR)

## Infectious Pustular Vulvovaginitis (IPV)

### *(Herpeviridae, alphaherpevirinae)*

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Conjunctival, vaginal, preputial scraping smear (FA)
  - Vesicle fluid, scraping, or swab (EM)
  - Nasopharyngeal, ocular, vaginal, preputial swab or fluid, semen (VI, PCR)
  - Paired serum (Serology—SN, ELISAb, IFA)
- Postmortem specimens
  - Lung, turbinate, trachea, salivary; fetal lung, liver, spleen, kidney, adrenal, cotyledon (FA, VI, PCR)
- Clinicopathology—respiratory, conjunctivitis, reproductive (dermal and abortifacient)
- BHV-1 causes two distinct clinical syndromes, a systemic respiratory infection (IBR), and dermal eruptions (pustules) of the genital mucosa (IPV), which also similarly affects bulls as Infectious Pustular Balanoposthitis.
- Abortion can result from IBR, typically not IPV infection.



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# Bovine Leukemia Virus (BLV)

## *(Retroviridae, deltaretrovirus)*

- Systemic infection / persistence / moderate significance (particularly Southern US)
- Antemortem specimens
  - Serum (Serology—AGID, ELISAb, federally approved for export testing)
  - Citrated blood (VI, PCR; difficult to culture)
- Postmortem specimens
  - Clotted blood, spleen (Serology)
  - Blood, lung, spleen, bone marrow (PCR, VI; difficult to culture)
- Clinicopathology—B-lymphocytosis & sarcoma
- Most infections are asymptomatic; about 30% of seropositive animals develop lymphocytosis with no other signs; only a few percent develop multisystemic tumors
- Sheep and goats are susceptible, but natural infection is rare
- Spread by contact with cellular transfer; possible iatrogenic transmission



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# Bovine Immunodeficiency Virus (BIV) (*Retroviridae, lentivirus*)

- Systemic infection / persistence / unknown significance (possibly moderate-major in Southern US)
- Antemortem specimens
  - Serum (Serology—ELISAb)
  - Whole blood (VI, PCR; viral detection not routine)
- Postmortem specimens
  - Clotted blood, spleen (Serology— ELISAb; not routine)
  - Blood, spleen, bone marrow, lymph node (effector lymphoid tissue), brain (VI, PCR, IHC, ISH; viral detection not routine)
- Clinicopathology (proposed) — acceleration and exacerbation of chronic problems from bacterial infections, neoplasia, and aging; lameness and neurological disease
- Seroprevalence in southern US cattle is far greater than northern cattle



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# Bovine Respiratory Syncytial Virus (BRSV) (*Paramyxoviridae, pneumovirus*)

- Local infection / no persistence / moderate-major significance
- Antemortem specimens
  - Nasopharyngeal, tracheal swab or fluid (FA, VI, PCR; culture not routine, difficult to culture)
  - Serum, paired (Serology—IFA, SN)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, VI, PCR; culture not routine)
- Clinicopathology—in adults, inapparent infection; in young calves, pneumonia, pulmonary edema, and emphysema, with secondary infections; "shipping fever" agent
- Protection from natural infection short-lived, re-infections common; current vaccine protocol minimally effective



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# Parainfluenza Type 3 (PI-3) Virus (*Paramyxoviridae, respirovirus*)

- Local infection / no persistence / moderate significance
- Antemortem specimens
  - Nasopharyngeal or tracheal swab or fluid (FA, VI, PCR)
  - Paired serum (Serology—IFA, HI, SN)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, VI, PCR)
- Clinicopathology—upper and lower respiratory disease with secondary infections; "shipping fever"
- Infectious for horses, dogs, primates, humans, and other ruminants



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# Bovine Respiratory Coronavirus (BRCV) (*Coronaviridae, coronavirus*)

- Local infection / unknown persistence / possible moderate significance
- Antemortem specimens
  - Nasopharyngeal or tracheal swab or fluid (FA, PCR, VI; culture not routine, requires specific cell line)
  - Serology: Paired serum (IFA, SN)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, PCR, VI; culture not routine)
- Clinicopathology—upper and lower respiratory disease, possible "shipping fever"



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# Miscellaneous Respiratory Viruses of Cattle

**Bovine Rhinovirus, three serotypes (*Picornaviridae, rhinovirus*)**  
**Bovine Adenovirus, ten serotypes (*Adenoviridae, mastadenovirus*)**

- Cause mild upper respiratory disease in cattle, and are of slight significance
- Sampling protocol is similar to PI-3 and BRSV in cattle



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# Mucosal Disease (MD)

## Bovine Viral Diarrhea (BVD) Virus

### *(Flaviviridae, pestivirus)*

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Nasopharyngeal swab or fluid, serum (ELISA<sub>Ag</sub> kit)
  - Nasopharyngeal swab or fluid, stool, citrated blood, serum (VI, PCR)
  - Paired serum (Serology—SN, ELISA<sub>Ab</sub>, IFA) (Carriers' serum is virus positive, antibody negative)
- Postmortem specimens
  - Turbinate, trachea, lung, tonsil, ileocecal Peyer's patches, mandibular and mesenteric lymph nodes, cerebellum (generally unrewarding samples include spleen, thymus, and mucosa of esophagus, rumen, and oral cavity) (FA, VI, PCR)
- Clinicopathology—mucosal disease, watery diarrhea, lameness, erosive stomatitis, pneumonia affecting only a few in a herd
- BVD (acute infection) and MD (chronic infection) are two distinct syndromes caused by the same virus. MD involves in-utero infection by non-cytopathic BVD, persistent infection in the calf, then mutation or super-infection by cytopathic BVD.
- Signs of MD are similar to BVD but more severe and of higher morbidity and mortality than acute BVD infection



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# Rotavirus (RotV)

## (*Reoviridae, rotavirus*)

- Local infection / no persistence / minor to major significance, especially in group habitats
- Antemortem specimens
  - Stool (serial, pooled) (EM, ELISA group specific kit)
- Postmortem specimens
  - Small intestine and contents, various levels (FA, EM)
- Clinicopathology—diarrhea, severe in neonates; mild to inapparent in older animals
- Found in most mammalian species; cross infections suspected
- Colostrum from vaccinated gravid dam confers immunity to suckling neonates in those species capable of passive transfer



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# Bovine Enteric Coronavirus (BECV) (*Coronaviridae, coronavirus*)

- Local infection / no persistence / minor significance
- Antemortem specimens
  - Stool (serial, pooled) (EM, VI ELISAg kit)
- Postmortem specimens
  - Intestine and contents (various levels) (FA, EM, VI)
- Clinicopathology—neonatal diarrhea, inapparent to subclinical in older animals



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# Bovine Viral Diarrhea (BVD) Virus

## *(Flaviviridae, pestivirus)*

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Dam's nasopharyngeal swab or fluid, citrated blood; carrier calf's serum (VI, PCR, ELISAg, commercial test kit)
  - Dam's paired serum (Serology—SN, ELISAb, IFA)
- Postmortem specimens
  - Fetal lung, liver, kidney, spleen, adrenal, brain (FA, VI, PCR)
- Clinicopathology—respiratory, enteric, dermal, mucosal disease, reproductive (early embryonic death, abortion, and persistently infected calves), cerebellar hypoplasia



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# Bovine Herpesvirus Type 1 (BHV-1), Infectious Bovine Rhinotracheitis (IBR) (*Herpeviridae, alphaherpevirinae*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Dam's conjunctival, vaginal scraping or smear (FA)
  - Dam's vesicle fluid, scraping, or swab (EM)
  - Dam's nasopharyngeal, ocular, vaginal swab or fluid (VI, PCR)
  - Dam's paired serum (Serology—SN, ELISAb, IFA)
- Postmortem specimens
  - Fetal lung, liver, spleen, kidney, salivary, adrenal, cotyledon (FA, VI, PCR)
- Clinicopathology—respiratory, conjunctivitis, reproductive (dermal and abortifacient)
- BHV-1 causes two distinct clinical syndromes, a systemic respiratory infection (IBR), and dermal eruptions (pustules) of the genital mucosa (IPV), which also similarly affects bulls in Infectious Pustular Balanoposthitis.
- Abortion can result from IBR, typically not IPV infection.



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# Bluetongue Virus (BTV)

## (*Reoviridae, orbivirus*)

- Systemic infection / prolonged viremia > 100 days / moderate significance in cattle as possible abortion agent
- Antemortem specimens
  - Dam's serum (Serology—AGID, ELISAb, federally approved testing for export)
  - Dam's citrated blood (VI, PCR)
- Postmortem specimens
  - Fetal lung, spleen, bone marrow (FA, VI, PCR) (BTV typically is not recovered from fetal tissues)
- Clinicopathology—asymptomatic or subclinical in non-gravid cattle; post-infection abortion in gravid cattle
- Replicates in hematopoietic and endothelial cells of blood vessels
- Mosquito vector (principally *Culicoides spp.* or "no-see-ems")
- Five serotypes (2, 10, 11, 13, 17) in North America



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# Bovine Mammillitis Virus (BHV-2) (*Herpesviridae, alphaherpevirinae*)

- Systemic infection / persistence / minor significance
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, including periphery) (EM, VI, PCR)
  - Serum (to discern carriers) (Serology—SN, IFA, ELISAb)
- Clinicopathology—ulcerating vesicles of teats and udder of first-calf heifers
- Differentiate from warts, cowpox, pseudocowpox, vesicular stomatitis, and foot and mouth disease



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# Bovine Papillomavirus

## *(Polyomaviridae, papillomavirus)*

- Systemic infection / persistence / minor significance
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, including periphery of lesions) (EM, PCR)
- Clinicopathology—cutaneous warts, fibropapillomas, squamous carcinoma
- Six types of Bovine papillomaviruses, distinguishable by the lesions they produce
  - Type 1—“teat frond” warts
  - Type 2—common cutaneous warts
  - Types 3 & 6—flat, broad lesions without fibroblast proliferation
  - Type 4—as 3 & 6, but with a tendency for alimentary and urinary squamous carcinoma
  - Type 5—“rice grain” fibropapillomas
- Horses contract infection by bovine types 1 & 2 from contaminated environment, and develop warts and sarcoids



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# Vesicular Stomatitis Virus (VSV) (*Rhabdoviridae, vesiculovirus*)

- Local infection / no persistence / minor significance
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, include periphery) (EM, VI, PCR)
- Clinicopathology—fever, vesicular lesions on tongue, oral mucosa, teats, coronary bands; epithelial denudation, secondary infections; resolves in 7 days
- Cross-species infection: horse, cow, pig, and deer
- Indiana and New Jersey strains are the two principal strains in North America; no cross protection
- VSV in cattle is reportable, requires federal program testing to distinguish from FMDV



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# Pseudocowpox Virus

## (*Poxviridae, parapoxvirus*)

- Local infection / no persistence / minor significance
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, include periphery) (EM, VI, PCR)
- Clinicopathology—papules on teats; scabs desquamate leaving characteristic ring or horseshoe lesions
- Zoonotic; “milker’s nodule”



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# Foot and Mouth Disease (FMD) Virus

## *(Picornaviridae, aphthovirus)*

- Systemic infection / persistence, prolonged shedding / not significant where eradicated; moderate-major where endemic
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, include periphery, buffered transport), probang collection of pharyngeal epithelium (VI, ELISAg, EM, PCR)
  - Serum (Serology—AGID, SN, ELISAb, CF)
- Clinicopathology—fever, vesicles on tongue, interdigital skin, oral mucosa, teats, coronary bands; ulcers, epithelial denudation, lameness, secondary infections
- Cross-species infection: cow, goat, pig, sheep, wild ruminants (not horse)
- Seven types (O, A, C, SAT-1, -2, -3, Asia-1) and numerous sub-types of varying antigenic composition preclude a universally efficacious vaccination program
- Eradicated in North and Central America, Australia, Japan, and most of Europe; controlled in South America; common in Asia, Africa, and elsewhere
- Reportable, subject of eradication program; **when suspected, contact state or federal animal health authorities**



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# Bluetongue Virus (BTV) (*Reovirus, orbivirus*)

- Systemic infection / prolonged viremia >100 days / major significance in sheep
- Antemortem specimens
  - Serum (Serology—AGID, ELISAb, federally approved testing for export)
  - Whole blood (VI, PCR)
- Postmortem specimens
  - Lung, spleen, bone marrow (FA, VI, PCR)
- Clinicopathology—in sheep, highly morbid systemic, hematogenous or hemorrhagic, dermal, lameness, abortion; in deer and antelope, hemorrhagic disease of high mortality; in cattle and goats, subclinical with abortion
- Replicates in hematopoietic and endothelial cells of blood vessels
- Mosquito vector (principally *Culicoides spp.* or "no-see-ems")
- Five serotypes (2, 10, 11, 13, 17) in North America



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# Epizootic Hemorrhagic Disease (EHD) Virus (*Reoviridae, orbivirus*)

- Systemic infection / prolonged viremia >100 days / moderate significance in deer
- Antemortem specimens
  - Serum (Serology—AGID)
  - Whole blood (VI, PCR)
- Postmortem specimens
  - Lung, spleen, bone marrow (FA, VI, PCR)
- Clinicopathology—hemorrhagic disease of high morbidity in deer; asymptomatic infection in non-gravid cattle, suspected post-infection abortion in gravid cattle
- Southern deer seem more resistant to disease, perhaps from a higher rate of immunity possibly due to the endemic state of EHD in the south
- Two serotypes in North America



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# Caprine Arthritis-encephalitis (CAE) Virus (*Retroviridae, lentivirus*)

- Systemic infection / persistence / major significance in goats
- Antemortem & postmortem specimens
  - Serum (Serology—AGID, ELISAb)
  - Synovial fluid or membrane (PCR, VI; culture not routine, requires specific cell line)
- Clinicopathology—arthritis in goats >12 months of age; encephalitis in kids 2-4 months of age; interstitial pneumonia; target organ is the mammary gland, with shedding; asymptomatic infections are believed to be common
- About 80% of all goats have antibody to CAEV
- Lab techniques not helpful in CAE diagnosis, which remains clinical and histological
- Serology testing is used in “test-and-slaughter” control programs



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# Ovine Progressive Pneumonia (OPP) Virus Maedi / Visna Virus (*Retroviridae, lentivirus*)

- Systemic infection / persistence / moderate significance in sheep
- Antemortem & postmortem specimens
  - Serum (Serology—AGID)
  - Viral detection not routinely performed
- Clinicopathology—insidious onset of pneumonia following protracted incubation (>2 years); often complicated by secondary infections
- Maedi is a synonym for respiratory disease caused by OPPV



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# Malignant Catarrhal Fever (MCF) Viruses (*Herpesviridae, gammaherpesvirus*)

- Systemic infection / persistence (latency) / slight (unknown) significance
- Antemortem specimens
  - Whole blood (peripheral leukocytes) (PCR)
  - Serum (ELISAb)
- Postmortem specimens
  - Lymph node, brain, gut, spleen (PCR)
- Clinicopathology—fever, depression, leukopenia, nasal & ocular discharge, ophthalmia, lymphadenopathy, diarrhea from intestinal mucosal erosions
- MCF virus group includes Alcelaphine Herpesvirus 1 (AHV-1, which causes classic MCF in African wildebeests), Ovine Herpesvirus type-2 (OHV-2, which crosses species to cause MCF in co-grazing domestic cattle or deer), and a putative goat gammaherpes (not yet associated with disease)
- MCF viruses can infect wild, native, or zoo ruminants, particularly cervids, which also have their own unique MCF viruses
- Except for AHV-1, the MCF viruses are uncultivable, and only are detected genetically
- Domestic MCF increasingly is recognized as a wildlife and zoo problem



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# Miscellaneous Respiratory Viruses of Sheep

**Ovine Adenovirus (*Adenoviridae, mastadenovirus*)**

**Respiratory Syncytial Virus (*Paramyxoviridae, pneumovirus*)**

- Have been associated with respiratory disease in sheep and goats, but are of slight clinical significance; sampling protocol is similar to BRSV in cattle

**Ovine Pulmonary Adenomatosis Virus (*Retroviridae, betaretrovirus*)**

- Causes a respiratory disease of moderate significance, worldwide distribution; diagnosis is cytological and histological



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# Coronavirus

## (*Coronaviridae, coronavirus*)

- Local infection / unknown persistence / moderate significance
- Antemortem specimens
  - Stool (serial, pooled) (EM)
- Postmortem specimens
  - Intestines and contents (EM)
- Clinicopathology—diarrhea, possibly severe in neonates; possibly subclinical-asymptomatic in older animals, with shedding



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# Miscellaneous Enteric Viruses

**Astrovirus (*Astroviridae, astrovirus*)**

**Adenovirus (*Adenoviridae, mastadenovirus*)**

- Local infection / no known persistence / slight significance
- Antemortem specimens
  - Stool (serial, pooled) (EM)
- Postmortem specimens
  - Intestines and contents (EM)
- Clinicopathology—diarrhea, possibly severe in neonates; possibly subclinical-asymptomatic in older animals, with shedding



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# Orf, Contagious Pustular Dermatitis (*Poxviridae, parapoxvirus*)

- Local infection / no persistence / minor significance
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, include periphery) (EM)
- Clinicopathology—cutaneous nodules that occasionally evolve into ulcers in areas where wool/hair is shortest, as head, neck, ears, axillae, under tail



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# Pseudorabies Virus (PRV)

## (*Herpesviridae, alphaherpesvirinae*)

- Systemic infection / persistence / moderate-major significance among ferals, eradicated from domestic stock in Louisiana
- Antemortem specimens
  - Single serum (Serology—SN, ELISAb, LA; federally approved testing, depends on state's vaccination program parameters)
  - Saliva, nasopharyngeal or tracheal swab/wash (VI, PCR)
- Postmortem specimens
  - Lung, trachea, tonsil, salivary, brain; fetal lung, spleen, liver, kidney, adrenal (FA, VI, PCR)
- Clinicopathology—respiratory, neurological, abortion
- Cross-species infection, neurological in secondary hosts: "mad-itc" or "pseudo-rabies" in cattle, feral pig hunting dogs



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# Porcine Reproductive and Respiratory Syndrome (PRRS) Virus, Lelystad Virus (*Arteriviridae, arterivirus*)

- Systemic infection / persistence / moderate-major significance
- Antemortem specimens
  - Paired serum (Serology—IFA, ELISAb)
  - Lung lavage (FA, IHC)
  - Nasopharyngeal, tracheal swab, wash, semen; blood or serum (VI, PCR; culture requires specific cell line)
- Postmortem specimens
  - Lung, lung lavage, spleen (FA, IHC, VI, PCR; culture requires PAM or MA-104 cell lines)
- Clinicopathology—abortion, respiratory in surviving neonates; subclinical in shedding breeders
- PRRSV best recovered from carrier sows; not detected in all individuals within affected litters



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# Porcine Hemagglutinating Encephalitis Virus (HEV) (*Coronaviridae, coronavirus*)

- Systemic infection / persistence / minor significance
- Antemortem specimens
  - Serum, herd profile (Serology—IFA, HI, SN)
  - Nasopharyngeal, tracheal swab or wash (PCR, VI; culture not routine)
- Postmortem specimens
  - Turbinate, trachea, lung, brain (FA, PCR, VI; culture not routine, requires primary pig cell culture)
- Clinicopathology—In pigs 2 weeks of age or younger, neurologically induced vomiting and wasting, rapidly progresses to general neurological signs; probably subclinical or asymptomatic in older pigs



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# Porcine Circovirus Type 2 (PCV-2) Postweaning Multisystemic Wasting Syndrome (PMWS) (*Circoviridae, circovirus*)

- Systemic infection / unknown persistence / unknown significance ("emerging disease")
- Antemortem specimens
  - Serum, herd profile (Serology—IFA, ELISAb; not routinely available)
  - Nasopharyngeal, tracheal swab or wash (PCR, VI; culture not routine)
- Postmortem specimens
  - Lung, inguinal lymph node, tonsil, or other lymphoid tissue, gut (rapid-growing tissues) (FA, PCR, VI; culture not routine)
- Clinicopathology—vomiting and wasting of piglets, interstitial pneumonia and enlarged lymph nodes, other pig viral diseases exacerbated by PCV-2 infection



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# Encephalomyelocarditis Virus (EMC) (*Picornaviridae, cardiovirus*)

- Systemic infection / no persistence / minor significance
- Antemortem specimens
  - Serum (Serology—SN, IFA, ELISAb; not routine)
  - Stool, blood (VI, PCR; not routine)
- Postmortem specimens
  - Heart, liver, spleen, kidney, pancreas, brain (VI, PCR; not routine)
- Clinicopathology—sudden death from myocarditis; few signs or gross lesions in pigs
- Rats are the primary host and reservoir of EMCV, which has a broad host range, including calves, horses, squirrels, swine, monkeys, elephants, and humans



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# Porcine Enterovirus Virus Type 1

## Porcine Polioencephalomyelitis (*Picornaviridae, enterovirus*)

- Systemic infection / no persistence / minor significance
- Antemortem specimens
  - Nasopharyngeal swab or wash, stool, (VI, PCR; not routine)
- Postmortem specimens
  - Brain, spinal cord (VI, PCR; not routine)
- Clinicopathology—reversible ataxia to 75% mortality; neurological signs include tremors, seizures, and paralysis, particularly in young pigs
- Synonyms—Teschen, talfan, endemic posterior paresis of pigs



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# Hog Cholera (HC)

## *(Flaviviridae, pestivirus)*

- Systemic infection / persistence / eradicated in North and Central America, but of major significance throughout underdeveloped areas of the world
- Antemortem specimens
  - Whole blood (buffy coat) (VI, PCR; isolates typically non-cytopathic)
  - Serum, herd profile (Serology—IFA, SN)
- Postmortem specimens
  - Lymph node (mandibular), tonsil, spleen, intestine (FA, VI, PCR)
- Clinicopathology—febrile, catarrhal, hemorrhagic disease of pigs affecting most tissue and organ systems
- HCV is antigenically and biologically similar to BVDV



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# Porcine Lymphosarcoma Virus (*Retroviridae, gammaretrovirus*)

- Systemic infection / persistent / minor significance in pigs
- Diagnosis by cytology and histopathology
- Clinicopathology—lymphocytosis



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# Swine Influenza Virus (SIV) (*Orthomyxoviridae, influenza A*)

- Local infection / no persistence / moderate significance
- Antemortem specimens
  - Nasopharyngeal swab or wash (VI, ELISAg kit, PCR)
  - Serum, herd profile (Serology—ELISAb, HI)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, VI, PCR)
- Clinicopathology—upper and lower respiratory, with fever, high morbidity, low mortality; H1N1 also pathogenic for humans and turkeys
- Prevalent strains, H3N2, H1N1; strains H1N2 and H4N6 also reportedly associated with pig respiratory disease



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# Swine Cytomegalovirus

## *(Retroviridae, betaherpesvirus)*

- Local infection / unknown persistence / minor significance in pig respiratory disease
- Sampling protocol is similar to BRSV in cattle



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# Transmissible Gastroenteritis (TGE) virus (*Coronaviridae, coronavirus*)

- Local infection / no persistence / minor-moderate significance
- Antemortem specimens
  - Stool (serial, pooled) (EM)
- Postmortem specimens
  - Small intestine and contents (FA, EM)
- Clinicopathology—neonatal diarrhea



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# Porcine Parvovirus (PPV) (*Parvoviridae, parvovirus*)

- Systemic infection / persistence / moderate significance
- Antemortem specimens
  - Dam's paired serum, herd profile (Serology—HI, SN)
- Postmortem specimens
  - Fetal lung, kidney (FA, PCR, VI; culture not routine)
  - Fetal body fluid, tissue pool (HAg vs. guinea pig RBCs)
- Clinicopathology—stillbirth, mummification, embryonic death, infertility (SMEDI)
- Difficult to recover from retained or mummified fetal material



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# Porcine Enteroviruses 2-11 (SMEDI group) (*Picornaviradae, enterovirus*)

- Systemic infection / unknown persistence / minor disease significance
- Antemortem specimens
  - Dam's serum (Serology—not routine, many serotypes)
  - Dam's stool (PCR, VI; culture not routine)
- Postmortem specimens
  - Fluids from fetus > 70 days gestation (Serology—SN)
  - Lung, brain, tonsil, kidney, spleen, intestinal contents (PCR, VI; culture not routine)
- Clinicopathology—stillbirth, mummification, embryonic death, infertility (SMEDI)
- Virus typically recoverable from stillborn tissues, but not mummified tissues



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# Swinepox Virus

## (*Poxviridae, suipoxvirus*)

- Local infection / no persistence / minor significance
- Antemortem & postmortem specimens
  - Vesicular fluid, active lesions (multiple samples, include periphery) (EM, VI, PCR)
- Clinicopathology—cutaneous nodules that occasionally evolve into ulcers



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# Miscellaneous Exanthematous Viruses of Swine

## Swine Vesicular Disease (*Enteroviridae, enterovirus*)

- Systemic infection / no persistence / minor significance in Asia and Europe only

## Vesicular Exanthema of Swine (*Caliciviridae, vesivirus*)

- Systemic infection / no persistence / no significance. Last seen in the US in 1959; believed to be eradicated from swine, but may still exist among certain marine mammals (i.e., sea lions); clinically indistinguishable from foot and mouth disease



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# Equine Infectious Anemia (EIA) Virus (*Retroviridae, lentivirus*)

- Systemic infection / persistence / moderate significance in the Southern US delta and coastal regions; slight-minor elsewhere
- Antemortem specimens
  - Serum (Serology—AGID, ELISAb, federally approved testing)
  - Whole blood (VI, PCR; viral detection not routine)
- Postmortem specimens
  - Serum (Serology—AGID, ELISAb)
  - Clotted blood, spleen (VI, PCR, AGID; viral detection not routine)
- Clinicopathology—"swamp fever" of equids; typically subclinical; primary infection febrile respiratory; chronic infection and shedding; eventual glomerulonephritis
- Mandatory testing and eradication programs, depending on federal and state regulations



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# Equine Viral Arteritis (EVA) Virus (*Arteriviridae, arterivirus*)

- Systemic infection / persistence / minor significance
- Antemortem specimens
  - Paired serum (Serology—modified SN, IFA, export testing requirements may apply)
  - Nasopharyngeal, conjunctival swab or wash, citrated blood, semen (PCR, VI; viral detection not routine)
- Postmortem specimens
  - Lung, turbinate, trachea, spleen, colon, cecum & associated lymph nodes, adrenal cortex; include small and medium-sized arteries (FA, PCR, VI; viral detection not routine)
- Clinicopathology—subclinical febrile illness with leukopenia, depression, edema, panvasculitis, edema; “abortion storms” on breeding farms
- Virus culturable only for first 2 weeks post-infection



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# Eastern Equine Encephalitis (EEE) Virus (*Togaviridae, alphavirus*)

- Systemic infection / no persistence / major significance in gulf delta and east coast US (swampy, wooded habitat), moderate-minor elsewhere
- Antemortem specimens
  - Serum, CSF (Serology—ELISAb for IgM, HI, SN)
  - Whole blood, brain biopsy (VI, PCR; low, variable viremia in horses, viral detection not routine)
- Postmortem specimens
  - Brain, spleen (FA, VI, PCR)
- Clinicopathology—EEE "sleeping sickness" of horses, rapid onset and progression of neurological signs, high mortality
- The horse, as most other mammalian species, is considered a dead-end host
- In EEE infections involving non-native birds as ratites, high viral titers are found in hematogenous organs, as the spleen
- Zoonotic, public health concern, reportable



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# West Nile Virus (WNV)

## *(Flaviviridae, flavivirus)*

- Systemic infection / no persistence / moderate-major significance
- Antemortem specimens
  - Serum, CSF (Serology—ELISAb for IgM; PRNT, HI, IFA, SN)
  - Whole blood, brain biopsy (VI, PCR; low, variable viremia in horses)
- Postmortem specimens
  - Brain (especially brain stem), spleen, kidney (VI, PCR, IHC)
- Clinicopathology—ataxia, limb weakness, recumbency, muscle fasciculation, fever, lip paralysis, and blindness; most neurological manifestations are associated with viral lesions in the brain stem; high mortality in clinically affected animals, but asymptomatic or subclinical infections are possible
- WNV ecology is supported by the bird-mosquito cycle, as EEE
- The horse, as most other mammalian species, is considered a dead-end host
- WNV surveillance efforts utilize kidney, heart, spleen, and brain tissue from sentinel avian species (crows, blue jays, and raptors)
- Zoonotic, public health concern, reportable



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# Other Viral Encephalitides of Horses

## Western equine encephalitis (WEE) (*Togaviridae, alphavirus*)

- Systemic infection / no persistence / slight significance in Louisiana and points east; moderate-major significance in Texas and points west
- Sampling and testing protocol is similar to EEE

## Venezuelan equine encephalitis (VEE) (*Togaviridae, alphavirus*)

- Systemic infection / no persistence / no disease significance in North America; major significance in South & Central America
- Sampling and testing protocol is similar to EEE



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# Equine Herpesvirus Type 1 (EHV-1) (*Herpesviridae, alphaherpesvirinae*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Paired serum (Serology—SN, IFA)
  - Nasopharyngeal swab or wash, citrated blood, CSF (VI, PCR)
- Postmortem specimens
  - Lung, trachea, turbinate (FA, VI, PCR)
- Clinicopathology—primary infection often respiratory; abortion usually in the last 4 months of gestation; occasionally neurological in surviving foals



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# Equine Herpesvirus Type-4 (EHV-4) Equine Rhinopneumonitis Virus (ERV) (*Herpesviridae, alphaherpesvirinae*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Paired serum (Serology—SN, IFA)
  - Conjunctival smear (FA)
  - Nasopharyngeal, tracheal, or bronchoalveolar swab or wash, citrated blood (FA, PCR, VI; culture difficult, not routine)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, PCR, VI; culture not routine)
- Clinicopathology—subclinical febrile respiratory illness in foals >2 months old
- EHV-4 closely related, yet genetically distinct from EHV-1



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# Equine Influenza Virus (EIV) (*Orthomyxoviridae, influenza virus A*)

- Local infection / no persistence / major significance
- Antemortem specimens
  - Nasopharyngeal swab or wash (VI, ELISAg kit, PCR)
  - Paired serum, herd profile (Serology—ELISAb, HI)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, VI, PCR)
- Clinicopathology—febrile respiratory illness, resolves in 2 weeks; seasonal and stress related
- Two main serovars: EIV-1=H7N7; EIV-2=H3N8



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# Miscellaneous Respiratory Viruses of Horses

Equine Herpesvirus Type 2 (*Herpesviridae, gammaherpesvirinae*)  
Equine Rhinovirus Type 2 (*Picornaviridae, genus unassigned*)  
Equine Adenovirus (*Adenoviridae, mastadenovirus*)

- Have been associated with mild respiratory illness of horses and are of slight-minor significance
- Sampling protocol is similar to EHV-1



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# Equine Herpesvirus Type-1 (EHV-1) Equine Abortion Virus (*Herpesviridae, alphaherpesvirinae*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Dam's paired serum (Serology—SN, IFA)
  - Dam's nasopharyngeal swab or wash, citrated blood, CSF (VI, PCR)
  - Dam's vaginal scraping or swab (FA, VI, PCR)
- Postmortem specimens
  - Fetal lung, spleen, liver, kidney, adrenal, brain, spinal cord; Dam's pharynx, CSF (FA, VI, PCR)
- Clinicopathology—abortion usually in the last 4 months of gestation; occasionally neurological in foals
- EHV-1 easier to culture than EHV-4



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# Miscellaneous Exanthematous Viruses of Horses

**Equine Papilloma Virus (*Papovaviridae, papillomavirus*)**

**Equine Sarcoid (*Papovaviridae, papillomavirus*)**

- Local infection / persistence / minor significance
- Antemortem & postmortem specimens
  - Lesions (warts or sarcoids) active, multiple samples (EM, PCR)
- Clinicopathology—warts and sarcoids, respectively
- Genetic evidence links sarcoids to bovine papillomavirus types 1 & 2, which are non-productive in lesions and unculturable from equine tissue



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# Canine Distemper Virus (CDV) (*Paramyxoviridae, morbillivirus*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Conjunctival smear, citrated blood smear, buffy coat smear, CSF (FA)
  - Paired serum, CSF (Serology—ELISAb kit, IFA, SN)
  - Nasopharyngeal swab or wash, citrated blood, CSF (PCR, VI; culture difficult, not routine,)
- Postmortem specimens
  - Lung, spleen, brain (cerebellum), bladder, tonsil, lymph node, thymus (FA, PCR, VI; culture not routine)
- Clinicopathology—multisystemic, respiratory, alimentary, hyperkeratotic, immunosuppressive, both acute and chronic neurological manifestations; secondary infections common
- Natural host range includes ferret, raccoon, mink, skunk, fox



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# Canine Parvovirus Virus (CPV) Type 2 (*Parvoviridae, parvovirus*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Stool (serial, pooled) (ELISAg kit, EM, HA, PCR, VI; culture difficult, not routine)
  - Paired serum (Serology—IFA, HI, SN)
- Postmortem specimens
  - Intestine (with contents) thymus, spleen, lung (FA, EM, PCR, VI; culture not routine)
- Clinicopathology—immunosuppressive, multisystemic, enteric; secondary infections common
- CPV-2 occurs in two subtypes
- Canine parvovirus type 1 (CPV-1) causes mild enteric and hemorrhagic disease with cardiac involvement in neonates, and is of slight significance



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# Canine Adenovirus Type 1 (CAAdV-1) Infectious Canine Hepatitis (ICH) Virus (*Adenoviridae, mastadenovirus*)

- Systemic infection / persistence / moderate significance
- Antemortem specimens
  - Paired serum (Serology—IFA, HI)
  - Nasopharyngeal or ocular swab or wash, citrated blood, urine (PCR, VI; culture not routine)
- Postmortem specimens
  - Liver, kidney, lung, bladder, spleen, lymph nodes, lung, brain (FA, PCR, VI; culture not routine)
- Clinicopathology—respiratory, alimentary, hepatitis; acute in pups; subclinical & chronic in older dogs; corneal opacity; encephalitis in foxes



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# Canine Adenovirus Type 2 (CAAdV-2)

## Canine Tracheobronchitis

### (*Adenoviridae, mastadenovirus*)

- Local infection / persistence / moderate significance
- Antemortem specimens
  - Paired serum (Serology—IFA, HI)
  - Nasopharyngeal or tracheal swab, wash or lavage (FA, PCR, VI; culture not routine)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, PCR, VI; culture not routine)
- Clinicopathology—"kennel-cough" respiratory complex; secondary infections



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# Canine Parainfluenza Virus (CPIV) Parainfluenza Virus Type 2 (PI-2) (*Paramyxoviridae, rubulavirus*)

- Local infection / no persistence / minor significance
- Antemortem specimens
  - Paired serum (Serology—IFA, HI)
  - Nasopharyngeal or tracheal swab, wash or lavage (FA, PCR, VI; culture not routine)
- Postmortem specimens
  - Turbinate, trachea, lung (FA, PCR, VI; culture not routine)
- Clinicopathology—"kennel-cough" respiratory complex, secondary infections common



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# Canine Herpesvirus (CHV)

## *(Herpesviridae, alphaherpesvirinae)*

- Local and systemic infection / persistence / slight significance as a respiratory pathogen, moderate significance in neonatal disease
- Antemortem specimens
  - Paired serum (pup, dam or adult) (Serology—IFA, SN, ELISAb)
  - Conjunctival, preputial, nasopharyngeal, or vaginal smear, swab, or wash (pup, dam, or adult) (FA, VI, PCR)
- Postmortem specimens
  - lung, kidney (neonate); lung, trachea, turbinate (juvenile or adult) (FA, VI, PCR)
- Clinicopathology—systemic infection of gravid dam results in hemorrhagic disease of neonates; local primary infection of juveniles and adults causes subclinical respiratory/reproductive tract disease



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# Feline Infectious Peritonitis (FIP) Virus (*Coronaviridae, coronavirus*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Serum, thoracic or abdominal fluid (Serology—ELISAb kit, IFA, serum or fluid protein electrophoresis)
  - Conjunctival scraping, smear, or swab, peritoneal lavage, needle biopsy, cavity fluid (FA, PCR; difficult to culture)
- Postmortem specimens
  - Kidney, liver, lung, parotid gland, thymus, lymph node (mesenteric), cecum, thoracic or abdominal fluid (FA, PCR; difficult to culture)
- Clinicopathology—wet or effusive (high protein fluid in body cavities); dry or noneffusive (pyogranulomatous lesions); ocular, neurological
- Finding of virus in intestine, but not in liver or kidney, suggests feline enteric coronavirus; conventional lab techniques do not readily distinguish FIP from other asymptomatic-subclinical coronavirus infections in cats



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# Feline Leukemia/sarcoma Virus (FeLV) (*Retroviridae, gammaretrovirus*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Serum, buccal swabs, saliva, citrated blood (ELISAg kit, PCR, VI; culture not routine)
  - Citrated blood smear, bone marrow biopsy smear (FA, indirect)
- Postmortem specimens
  - Bone marrow (ELISAg, FA indirect, PCR, VI; culture not routine)
- Clinicopathology—multisystemic, myeloproliferative, lymphosarcoma, fibrosarcoma, anemia



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# Feline Immunodeficiency Virus (FIV) (*Retroviridae, lentivirus*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Serum (Serology—ELISAb kit, IFA)
  - Whole blood, buccal swab, saliva (PCR, VI; viral detection not routine)
- Postmortem specimens
  - Serum from heart clot (Serology—ELISAb, IFA)
  - Effector lymphoid tissue (PCR, VI; viral detection not routine)
- Clinicopathology—enteric, dermal, neurological, general reticuloendothelial in older, mostly male cats; insidious onset of FUO, leukopenia, lymphadenopathy, anemia, secondary infections, unkempt and wasting; resembles various stages of human AIDS



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# Feline Parvovirus (FPV)

## Feline Panleukopenia Virus (FPL)

*(Parvoviridae, parvovirus)*

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Serum (Serology—ELISAb, IFA)
  - Stool (serial, pooled) (ELISAg, EM)
  - Urine, saliva (PCR, VI; difficult to culture)
- Postmortem specimens
  - Intestine, lung, lymph node, spleen, thymus (FA, EM, PCR, VI; difficult to culture)
- Clinicopathology—multisystemic, enteric, neonatal infection and disease; cerebellar hypoplasia and wasting disease in kittens



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# Feline Herpesvirus (FHV)

## Feline Viral Rhinotracheitis (FVR)

*(Herpesviridae, alphaherpesvirinae)*

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Conjunctival smear (FA)
  - Nasopharyngeal or conjunctival swab or wash (VI, PCR)
  - Paired serum (Serology—SN, IFA, ELISAb)
- Postmortem specimens
  - Lung, trachea, turbinate, spleen, tonsil (FA, VI, PCR)
- Clinicopathology—respiratory disease, acute in kittens; inapparent or subclinical in adults, conjunctivitis, post-infection abortion



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# Feline Calicivirus (FCV) (*Caliciviridae, vesivirus*)

- Systemic infection / persistence / major significance
- Antemortem specimens
  - Nasopharyngeal, conjunctival, sinus, oral, buccal swab or wash (VI, PCR)
  - Paired serum (Serology—SN, IFA, ELISAb)
- Postmortem specimens
  - Lung, trachea, turbinate, oral lesions (FA, VI, PCR)
- Clinicopathology—oral vesicles, upper respiratory disease
- Common secondary infection in FIV-infected cats



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# Feline Enteric Coronavirus (FECV) (*Coronaviridae, coronavirus*)

- Local infection / unknown persistence / minor significance
- Antemortem specimens
  - Stool (serial, pooled) (EM)
- Postmortem specimens
  - Intestine and contents (FA, EM, PCR, VI; culture not routine)
- Clinicopathology—mild to severe diarrhea in neonates; subclinical-asymptomatic with shedding in adults
- Conventional technology does not distinguish FECV from FIP
- Certain strains of FECV penetrate the intestines and migrate to the regional lymph nodes, where they theoretically mutate into FIP strains



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# Rabiesvirus

## (*Rhabdoviridae, lyssavirus*)

- Systemic infection / persistence / minor significance as an animal disease, major public health significance
- Antemortem specimens (antemortem diagnostic testing not routine)
  - Serum (Serology—IFA, ELISAb; immune status check, int'l travel)
  - Buccal or saliva swab, mussel or tactile hair, muscle skin biopsy (IHC, PCR, VI; antemortem viral detection not routine)
- Postmortem specimens
  - Brain (bilateral sample, including hippocampus, cerebellum, brain stem) (FA, VI, PCR)
- Clinicopathology—chronic neurological with shedding in reservoir hosts; progressive neurological in incidental hosts
- In Louisiana, bat and skunk are reservoir hosts; dog, cat, fox, raccoon, cattle, and horse are incidental hosts; all mammals are potential hosts
- Zoonotic, reportable



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- Virus has slight or no real clinical significance, or currently is not recognized to exist in North America.
- Sampling protocol is similar to that of other viruses with the same systemic involvement in this animal species



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